

# Package: GeneNMF (via r-universe)

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**Type** Package

**Title** Non-Negative Matrix Factorization for Single-Cell Omics

**Version** 0.6.0

**Description** A collection of methods to extract gene programs from single-cell gene expression data using non-negative matrix factorization (NMF). 'GeneNMF' contains functions to directly interact with the 'Seurat' toolkit and derive interpretable gene program signatures.

**Depends** R (>= 4.3.0)

**Imports** RcppML, Matrix, stats, methods, utils, Seurat (>= 4.3.0), cluster, lsa, irlba, pheatmap, viridis

**Suggests** knitr, rmarkdown, fgsea, msigdb

**VignetteBuilder** knitr

**URL** <https://github.com/carmonalab/GeneNMF>

**BugReports** <https://github.com/carmonalab/GeneNMF/issues>

**License** GPL-3

**Encoding** UTF-8

**LazyData** true

**RoxygenNote** 7.3.2

**Repository** <https://carmonalab.r-universe.dev>

**RemoteUrl** <https://github.com/carmonalab/genenmf>

**RemoteRef** HEAD

**RemoteSha** f7792fa15d7921e67a5e1e94e20ac149291b3772

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findVariableFeatures\_wfilters  
*Find variable features*

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### Description

Select highly variable genes (HVG) from an expression matrix. Genes from a blocklist (e.g. cell cycling genes, mitochondrial genes) can be excluded from the list of variable genes, as well as genes with very low or very high average expression

### Usage

```
findVariableFeatures_wfilters(
  obj,
  nfeatures = 2000,
  genesBlockList = NULL,
  min.exp = 0.01,
  max.exp = 3
)
```

### Arguments

obj	A Seurat object containing an expression matrix
nfeatures	Number of top HVG to be returned
genesBlockList	Optionally takes a vector or list of vectors of gene names. These genes will be ignored for HVG detection. This is useful to mitigate effect of genes associated with technical artifacts or batch effects (e.g. mitochondrial, heat-shock response). If set to 'NULL' no genes will be excluded
min.exp	Minimum average normalized expression for HVG. If lower, the gene will be excluded
max.exp	Maximum average normalized expression for HVG. If higher, the gene will be excluded

### Value

Returns the input Seurat object obj with the calculated highly variable features accessible through VariableFeatures(obj)

**Examples**

```
data(sampleObj)
sampleObj <- findVariableFeatures_wfilters(sampleObj, nfeatures=100)
```

---

getDataMatrix	<i>Extract data matrix from Seurat object</i>
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**Description**

Get the gene expression matrix from a Seurat object, optionally centered and/or subset on highly variable genes

**Usage**

```
getDataMatrix(  
  obj,  
  assay = "RNA",  
  slot = "data",  
  hvg = NULL,  
  center = FALSE,  
  scale = FALSE,  
  non_negative = TRUE  
)
```

**Arguments**

obj	Seurat object
assay	Get data matrix from this assay
slot	Get data matrix from this slot (=layer)
hvg	List of variable genes to subset the matrix. If NULL, uses all genes
center	Whether to center the data matrix
scale	Whether to scale the data matrix
non_negative	Enforce non-negative values for NMF

**Value**

Returns a sparse data matrix (cells per genes), subset according to the given parameters

**Examples**

```
data(sampleObj)
matrix <- getDataMatrix(sampleObj)
```

---

getMetaPrograms      *Extract consensus gene programs (meta-programs)*

---

### Description

Run it over a list of NMF models obtained using [multiNMF](#); it will determine gene programs that are consistently observed across samples and values of  $k$ .

### Usage

```
getMetaPrograms(
  nmf.res,
  nMP = 10,
  specificity.weight = 5,
  weight.explained = 0.5,
  max.genes = 200,
  metric = c("cosine", "jaccard"),
  hclust.method = "ward.D2",
  min.confidence = 0.5,
  remove.empty = TRUE
)
```

### Arguments

nmf.res	A list of NMF models obtained from <a href="#">multiNMF</a>
nMP	Total number of meta-programs
specificity.weight	A parameter controlling how specific gene should be for each program. ‘specificity.weight=0’ no constraint on specificity, and positive values impose increasing specificity.
weight.explained	Fraction of NMF weights explained by selected genes. For example if weight.explained=0.5, all genes that together account for 50% of NMF weights are used to return program signatures.
max.genes	Max number of genes for each programs
metric	Metric to calculate pairwise similarity between programs
hclust.method	Method to build similarity tree between individual programs
min.confidence	Percentage of programs in which a gene is seen (out of programs in the corresponding program tree branch/cluster), to be retained in the consensus metaprograms
remove.empty	Whether to remove meta-programs with no genes above confidence threshold

**Value**

Returns a list with i) 'metaprograms.genes' top genes for each meta-program; ii) 'metaprograms.metrics' dataframe with meta-programs statistics: a) freq. of samples where the MP is present, b) average silhouette width, c) mean similarity (cosine or Jaccard), d) number of genes in MP, e) number of gene programs in MP; iii) 'programs.similarity': matrix of similarities (Jaccard or cosine) between meta-programs; iv) 'programs.tree': hierarchical clustering of meta-programs (hclust tree); v) 'programs.clusters': meta-program identity for each program

**Examples**

```
library(Seurat)
data(sampleObj)
geneNMF_programs <- multiNMF(list(sampleObj), k=5)
geneNMF_metaprograms <- getMetaPrograms(geneNMF_programs, nMP=3)
```

---

getNMFgenes

*Get list of genes for each NMF program*


---

**Description**

Run it over a list of NMF models obtained using multiNMF()

**Usage**

```
getNMFgenes(
  nmf.res,
  specificity.weight = 5,
  weight.explained = 0.5,
  max.genes = 200
)
```

**Arguments**

nmf.res	A list of NMF models obtained using multiNMF()
specificity.weight	A parameter controlling how specific gene should be for each program. 'specificity.weight=0' no constraint on specificity, and positive values impose increasing specificity.
weight.explained	Fraction of NMF weights explained by selected genes. For example if weight.explained=0.5, all genes that together account for 50% of NMF weights are used to return program signatures.
max.genes	Max number of genes for each program

**Value**

Returns a list of top genes for each gene program found by multiNMF()

**Examples**

```
library(Seurat)
data(sampleObj)
geneNMF_programs <- multiNMF(list(sampleObj), k=5)
geneNMF_genes <- getNMFgenes(geneNMF_programs)
```

---

multiNMF

*Run NMF on a list of Seurat objects*


---

**Description**

Given a list of Seurat objects, run non-negative matrix factorization on each sample individually, over a range of target NMF components (k).

**Usage**

```
multiNMF(
  obj.list,
  assay = "RNA",
  slot = "data",
  k = 5:6,
  hvg = NULL,
  nfeatures = 2000,
  L1 = c(0, 0),
  min.exp = 0.01,
  max.exp = 3,
  center = FALSE,
  scale = FALSE,
  min.cells.per.sample = 10,
  hvg.blocklist = NULL,
  seed = 123
)
```

**Arguments**

obj.list	A list of Seurat objects
assay	Get data matrix from this assay
slot	Get data matrix from this slot (=layer)
k	Number of target components for NMF (can be a vector)
hvg	List of pre-calculated variable genes to subset the matrix. If hvg=NULL it calculates them automatically
nfeatures	Number of HVG, if calculate_hvg=TRUE
L1	L1 regularization term for NMF
min.exp	Minimum average log-expression value for retaining genes

max.exp	Maximum average log-expression value for retaining genes
center	Whether to center the data matrix
scale	Whether to scale the data matrix
min.cells.per.sample	Minimum number of cells per sample (smaller samples will be ignored)
hvg.blocklist	Optionally takes a vector or list of vectors of gene names. These genes will be ignored for HVG detection. This is useful to mitigate effect of genes associated with technical artifacts and batch effects (e.g. mitochondrial), and to exclude TCR and BCR adaptive immune(clone-specific) receptors. If set to 'NULL' no genes will be excluded
seed	Random seed

**Value**

Returns a list of NMF programs, one for each sample and for each value of 'k'. The format of each program in the list follows the structure of `nmf` factorization models.

**Examples**

```
library(Seurat)
data(sampleObj)
geneNMF_programs <- multiNMF(list(sampleObj), k=5)
```

---

multiPCA

*Run PCA on a list of Seurat objects*


---

**Description**

Given a list of Seurat objects, run non-negative PCA factorization on each sample individually.

**Usage**

```
multiPCA(
  obj.list,
  assay = "RNA",
  slot = "data",
  k = 4:5,
  hvg = NULL,
  nfeatures = 500,
  min.exp = 0.01,
  max.exp = 3,
  min.cells.per.sample = 10,
  center = FALSE,
  scale = FALSE,
  hvg.blocklist = NULL,
  seed = 123
)
```

**Arguments**

obj.list	A list of Seurat objects
assay	Get data matrix from this assay
slot	Get data matrix from this slot (=layer)
k	Number of target components for PCA
hvg	List of pre-calculated variable genes to subset the matrix. If hvg=NULL it calculates them automatically
nfeatures	Number of HVG, if calculate_hvg=TRUE
min.exp	Minimum average log-expression value for retaining genes
max.exp	Maximum average log-expression value for retaining genes
min.cells.per.sample	Minimum number of cells per sample (smaller samples will be ignored)
center	Whether to center the data matrix
scale	Whether to scale the data matrix
hvg.blocklist	Optionally takes a vector or list of vectors of gene names. These genes will be ignored for HVG detection. This is useful to mitigate effect of genes associated with technical artifacts and batch effects (e.g. mitochondrial), and to exclude TCR and BCR adaptive immune(clone-specific) receptors. If set to 'NULL' no genes will be excluded
seed	Random seed

**Value**

Returns a list of non-negative PCA programs, one for each sample. The format of each program in the list follows the structure of [nmf](#) factorization models.

**Examples**

```
library(Seurat)
data(sampleObj)
geneNMF_programs <- multiPCA(list(sampleObj), k=5)
```

---

plotMetaPrograms

*Visualizations for meta-programs*


---

**Description**

Generates a clustered heatmap for meta-program similarities (by Jaccard index or Cosine similarity). This function is intended to be run on the object generated by [getMetaPrograms](#), which contains a pre-calculated tree of pairwise similarities between clusters (as a 'hclust' object).



## Usage

```
plotMetaPrograms(  
  mp.res,  
  similarity.cutoff = c(0, 1),  
  scale = "none",  
  palette = viridis(100, option = "A", direction = -1),  
  annotation_colors = NULL,  
  main = "Clustered Heatmap",  
  show_rownames = FALSE,  
  show_colnames = FALSE,  
  ...  
)
```

## Arguments

mp.res	The meta-programs object generated by <a href="#">getMetaPrograms</a>
similarity.cutoff	Min and max values for similarity metric
scale	Heatmap rescaling (passed to pheatmap as 'scale')
palette	Heatmap color palette (passed to pheatmap as 'color')
annotation_colors	Color palette for MP annotations
main	Heatmap title
show_rownames	Whether to display individual program names as rows
show_colnames	Whether to display individual program names as cols
...	Additional parameters for pheatmap

## Value

Returns a clustered heatmap of MP similarities, in ggplot2 format

## Examples

```
library(Seurat)  
data(sampleObj)  
geneNMF_programs <- multiNMF(list(sampleObj), k=5)  
geneNMF_metaprograms <- getMetaPrograms(geneNMF_programs, nMP=3)  
plotMetaPrograms(geneNMF_metaprograms)
```

---

`runGSEA`*Run Gene set enrichment analysis*

---

### Description

Utility function to run Gene Set Enrichment Analysis (GSEA) against gene sets from MSigDB. Note: this is an optional function, which is conditional to the installation of suggested packages `fgsea` and `msigdb`.

### Usage

```
runGSEA(  
  genes,  
  universe = NULL,  
  category = "H",  
  subcategory = NULL,  
  species = "Homo sapiens",  
  pval.thr = 0.05  
)
```

### Arguments

<code>genes</code>	A vector of genes
<code>universe</code>	Background universe of gene symbols (passed on to <code>fgsea::fora</code> )
<code>category</code>	GSEA main category (e.g. "H" or "C5")
<code>subcategory</code>	GSEA subcategory
<code>species</code>	Species for GSEA analysis. For a list of the available species, type <code>msigdb::msigdb_species()</code>
<code>pval.thr</code>	Min p-value to include results

### Value

Returns a table of enriched gene programs from GSEA

### Examples

```
data(sampleObj)  
geneset <- c("BANK1", "CD22", "CD79A", "CD19", "IGHD", "IGHG3", "IGHM")  
#test is conditional on availability of suggested packages  
if (requireNamespace("fgsea", quietly=TRUE) &  
    requireNamespace("msigdb", quietly=TRUE)) {  
  gsea_res <- runGSEA(geneset,  
    universe=rownames(sampleObj),  
    category = "C8")  
}
```

---

`runNMF`*Compute NMF as a low-dim embedding for Seurat*

---

**Description**

Compute NMF embeddings for single-cell dataset, and store them in the Seurat data structure. They can be used as an alternative to PCA for downstream analyses.

**Usage**

```
runNMF(  
  obj,  
  assay = "RNA",  
  slot = "data",  
  k = 10,  
  new.reduction = "NMF",  
  seed = 123,  
  L1 = c(0, 0),  
  hvg = NULL,  
  center = FALSE,  
  scale = FALSE  
)
```

**Arguments**

<code>obj</code>	A seurat object
<code>assay</code>	Get data matrix from this assay
<code>slot</code>	Get data matrix from this slot (=layer)
<code>k</code>	Number of components for low-dim representation
<code>new.reduction</code>	Name of new dimensionality reduction
<code>seed</code>	Random seed
<code>L1</code>	L1 regularization term for NMF
<code>hvg</code>	Which genes to use for the reduction
<code>center</code>	Whether to center the data matrix
<code>scale</code>	Whether to scale the data matrix

**Value**

Returns a Seurat object with a new dimensionality reduction (NMF)

**Examples**

```
data(sampleObj)  
sampleObj <- runNMF(sampleObj, k=8)
```

---

`sampleObj`*Sample dataset to test GeneNMF installation*

---

**Description**

A Seurat object containing single-cell transcriptomes (scRNA-seq) for 50 cells and 20729 genes. Single-cell UMI counts were normalized using a standard log-normalization: counts for each cell were divided by the total counts for that cell and multiplied by 10,000, then natural-log transformed using 'log1p'.

This is a subsample of 25 predicted B cells and 25 predicted NK cells from the large scRNA-seq PBMC dataset published by Hao et al. ([doi:10.1016/j.cell.2021.04.048](https://doi.org/10.1016/j.cell.2021.04.048)) and available as UMI counts at [https://atlas.fredhutch.org/data/nygc/multimodal/pbmc\\_multimodal.h5seurat](https://atlas.fredhutch.org/data/nygc/multimodal/pbmc_multimodal.h5seurat)

**Usage**`sampleObj`**Format**

A sparse matrix of 50 cells and 20729 genes.

**Source**

[doi:10.1016/j.cell.2021.04.048](https://doi.org/10.1016/j.cell.2021.04.048)

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